

Molecular Detection Kits for GMO Soybean Meal QuickGuide

For Technical Support Call (207) 797-0300 Press 2

ENVIROLOGIX

Doc. M213-1215

1. Prepare Sample using Sample Extraction Set 5

- Pre-heat dry block to 95°C for at least 30 minutes Ι.
- Grind a representative sample in an Oster blender for 20 seconds 2.
- 3. Shake MB9 bottle for 5 seconds to distribute particulate matter; add I mL of MB9 to a 2 mL clear tube supplied with Extraction Set 5
- 4. Using a clean grain scoop, add four unpacked level scoops of ground sample to clear tube and cap tightly; shake tube to mix
- 5. Heat tube at 95°C for 6 minutes (±30 seconds)
- Remove tube and vortex for 5 seconds; sample may not mix completely 6.
- 7. Centrifuge sample at 10,000 x g for 3 minutes (±30 seconds)
- Add 100 µL of D4 buffer to a 1.5 mL blue tube supplied with Extraction Set 8.
- 9. Add 100 μ L of centrifuged sample to blue tube, avoiding particulates, and vortex to mix
- 10. 25 μ L of this sample will be used in the assay

2. Prepare for Testing

3. AmpliFire Setup

- Turn on AmpliFire Unit and allow to warm to 56°C Ι.
- 2. Ensure EnviroLogix GMO kit is at room temperature



Next

Star

Cancel

After warmup, Select " Execute Reaction"		56.0°C Reaction 1
Select " Scan Product Code " and scan barcode on Master Mix foil pouch using barcode camera on left side of reader	1 Execute Reaction View Results	Step FrClock "Sear" partners: Sco actar Of Code
Kit Identifier # will display		Cancel
Select " Next "		
Enter Run-Specific "Reaction Description" here	S6.0° C	Restort of a second sec
Enter Custom Fields if applicable	Stag 2: Enter a num an the Vescence Descaption Field	Contract of
Select Next	Step 2 Criter a cancer Midd wave is "Concerning of a cancer field 1 "Concerning" and the cancer field 2.8 Cancer field 2.8 Cancer field 2.8	

(1)

Cancel

Enter Well-Specific info one well at a • time or "Finish" to skip

•

•

.

.

•

•

Select Start and AmpliFire will be . ready to read strip tubes

Remove green Reaction Buffer strip tubes from the kit. Flick down prior to uncapping so that buffer is at bottom of tubes. Mark the left end for orientation.

- Transfer **25 µL of each sample** to the green Reaction Buffer vials. Use a fresh pipette tip for each transfer.
- **Recap tubes** and tap down or centrifuge to ensure all liquid is at the bottom of the tube
 - Using multi-channel pipette, transfer
 50 µL to clear Master Mix tubes.
 - Do **not** mix within the clear tube.



•

- Recap with Flat Caps (seal completely!!).
- Mark left side of cap for orientation.
- Flick and tap down several times to remove air bubbles. **No bubbles** in bottom of tubes!
- Place capped and inspected clear strip tube in instrument and immediately press "OK"
- After 15 minutes, results will be displayed as Not Detected (-) or Positive (+).
- After assay is complete, **remove run reaction strip tubes** discard in opened foil pouch. **Never open a completed reaction tube!**
- Export results from the Home Page (View Results/Export to USB)
- Clean up work area.



- Oster blender
- Precision pipettes to deliver 25-1000 μL (p-100 & p-1000)
- Pipette tips
- Dry heat block capable of 95±1°C (with insert capable of holding 2 mL tubes)
- Vortex
- Microcentrifuge capable of 10,000 x g
- Timer













- 2 -