

# QuickStix™Kit for DMO Bulk Soybeans



#### Highlights:

- Results in 5 minutes or less
- Three bulk soybean kits utilize Common Extraction<sup>TM</sup>:
  - Roundup Ready®
  - LibertyLink®
  - DMO
- Available as 100-strip kit, in bulk packaging, or in QuickCombs™

#### Contents of Kit:

- 100 QuickStix Strips packed in two moisture-resistant canisters
- 100 transfer pipettes
- 100 three-ounce cups

#### Items Not Provided:

 Blender for sample prep:

 Oster® Sunbeam blender model #4094 (with 4 oz. polystyrene blender jar, ice crusher blade, gasket, and

blender base) ~or~

- 2. Waring blender model
  31BL91 or equivalent (with
  glass Mason jars and jar
  adapter [Eberbach #E8495]
  along with protective cover
  ~or~
- 3. BUNN coffee grinder (industrial style grinder set on AutoDrip setting)
- Graduated cylinder
- Tap water

Catalog Number AS 050 BGB

Part #11770

### **Intended Use**

The EnviroLogix QuickStix Kit for DMO (Dicamba Mono-oxygenase) Bulk Soybeans is designed to extract and detect the presence of DMO protein at the levels typically expressed in soybean event MON 87708. The sensitivity of these QuickStix Strips is 0.25% DMO soybean (i.e. one DMO-expressing soybean in 400 non-DMO-expressing soybeans). For DMO detection in soy leaf and single seed samples, please use Cat. No. AS 050 LS.

#### **How the Test Works**

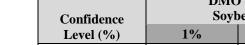
In order to detect the DMO protein expressed by event MON 87708 soybeans, the sample must first be extracted to solubilize the protein. Each QuickStix Strip has an absorbent pad at each end. The protective tape with the arrow indicates the end of the strip to insert into the reaction cup. The sample will travel up the membrane strip and be absorbed into the larger pad at the top of the strip. The portion of the strip between the protective tape and the absorbent pad at the top of the strip is used to view the reactions as described under "Interpreting the Results." Please avoid bending the strips.

### **Sample Preparation**

#### **Step 1: Determine Number and Size of Sub-samples**

- Collect a composite sample according to USDA/ GIPSA instructions found in the following reference documents:
- http://www.gipsa.usda.gov/fgis/handbook/gihbk1\_inspec.aspx USDA Grain Inspection Handbook, Book 1, Grain Sampling.
- http://www.gipsa.usda.gov/fgis/biotech/sample2.htm Guidance document entitled Sampling for the Detection of Biotech Grains.
- http://www.gipsa.usda.gov/fgis/biotech/sample1.htm Practical Application of Sampling for the Detection of Biotech Grains.
- www.gipsa.usda.gov/fgis/biotech/samplingplan1.xls This website provides a simple to use Sample Planner (29K Excel Spreadsheet). The planner allows you to enter different assumptions in terms of sample size, number of samples, acceptable quality level and to determine the probability of accepting lots with given concentration levels. It also plots the probabilities in graph form for easy interpretation. Specific data can be saved for documentation and future analyses.
- The following is a helpful reference for use in designing a sampling plan: Remund, K.M., Dixon, D.A., Wright D.L., Holden, L.R. "Statistical considerations in seed purity testing for transgenic traits", Seed Science Research, June 2001, Vol. 11 No.2, pp. 101-119.
- To select the appropriate sample size first determine the purity standard and the
  degree of confidence required. Confidence level means the statistical probability
  that the true DMO soybean level in the sample is below the selected purity
  standard.
- Table 1 provides a guideline for determining the number of 400 bean subsamples necessary to provide effective screening for different GM concentrations at a 95% confidence level.





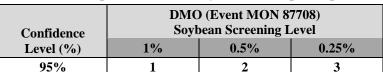


Table 1 - Number of representative 400-soybean subsamples required for testing



Choose grinding method

Soybean Common Extraction Grams of beans  $\times$  5 = mL water For example:  $(100 \times 0.15) = 15g$  $15g \times 5 = 75$  mL water

Extract in tap water



Fill vial to ridge with extract

#### Step 2: Determine Sub-sample Weight, Jar Size and Grind Times

- Determine average weight of individual bean lot to be tested (weigh 100 beans, divide by 100).
- Calculate the weight of the number of beans to sub-sampled and tested (number of beans X Average Weight/bean).
- Choose an appropriate jar size and grind time based on the type of blender available for sub-sample preparation (see Table 2). Oster Sunbeam Blender with ice crusher blade is recommended over the Waring Blender for its bean grinding efficiency. (Note that bean grind time is longer and requires additional steps\* when using a Waring Blender).

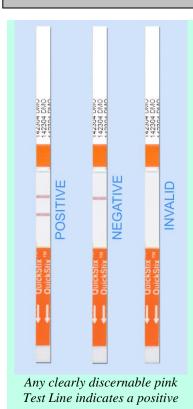
Table 2 - Soybeans

# of Beans		Sub-sample		Grind Time on High
(approximate)	Blender Type	weight (g)	Jar size (oz.)	Speed
100-200	Oster Sunbeam	16-38	8	20 seconds
100	Waring	16-38	8	60 seconds (2 X 30 sec.*)
200-400	Waring	38-65	16	60 seconds (2 X 30 sec.*)

For best results blend beans for ½ of total time, remove the jar and shake to redistribute larger particles, replace and resume grinding.

#### Prepare the sample

- Weigh sample into the appropriate vessel.
- Put protective cover over glass jars.
- Grind sample on high speed until all whole beans are finely ground.
- Add the volume of tap water calculated by the formula (above, left).
- Cap and shake jar vigorously until the entire sample is wet (20-30 seconds, depending on the number of grains). Sample will begin to settle immediately and liquid can be drawn off at that time. Avoid pulling up particles with the transfer pipette.
- Draw up liquid portion from above the settled sample and dispense extract into reaction vial until it is filled (this will take 2-3 transfers). Avoid pulling up particles. Allow extract to settle in the reaction vial for 30 seconds before adding a test strip.
- To prevent cross-contamination, thoroughly clean blender parts and jars of dust and residue prior to preparation of a second sample. Use a new transfer pipette and cup for each sample.



### How to Run the QuickStix Strip Test

- Allow refrigerated canisters to come to room temperature before opening.
   Remove the QuickStix Strips to be used. Avoid bending the strips. Reseal the canister immediately.
- Place the strip into the reaction vial. The sample will travel up the strip.
   Reaction vials will stand on their own or may be inserted into the cardboard racks provided.
- Allow the strip to develop for 5 minutes before making final assay interpretations. Positive sample results may become obvious much more quickly.
- To retain the strip, cut off and discard the bottom section of the strip covered by the arrow tape.

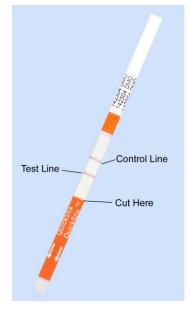
NOTE: Use extreme caution to prevent sample-to-sample cross-contamination with grain, fluids, or disposables.

## **Interpreting the Results**

Development of the Control Line within 5 minutes indicates that the strip has functioned properly. Any strip that does not develop a Control Line should be discarded, and the sample re-tested using another strip.

If the extract is from a sample containing at least 0.25% DMO soybean (1 in 400), a second line (Test Line) will develop on the membrane strip between the Control Line and the protective arrow tape. The results should be interpreted as positive for DMO protein expression.

If the extract is from a negative sample, the strip will only show the control line.



### Kit Storage

QuickStix can be stored at room temperature, or refrigerated for a longer shelf life. Note the shelf life on the kit box for each storage temperature. The kit may be used in field applications; however, prolonged exposure to high temperatures may adversely affect the test results. Do not open the desiccated canister until ready to use the test strips.

### **Precautions and Notes**

- This kit is designed to screen for presence or absence only, and is not meant to be quantitative.
- This product is currently only applicable for use in soybean bulk grain testing, and not applicable for use in any other crop or in soybean single seed or leaf testing.
- As with all tests, it is recommended that results be confirmed by an alternate method when necessary.







- The assay has been optimized to be used with the protocol provided in the kit. Deviation from this protocol may invalidate the results of the test.
- The results generated through the proper use of this diagnostic tool reflect the condition of the working sample directly tested. Extrapolation as to the condition of the originating lot, from which the working sample was derived, should be based on sound sampling procedures and statistical calculations which address random sampling effects, non-random seed lot sampling effects and assay system uncertainty. A negative result obtained when properly testing the working sample does not necessarily mean the originating lot is entirely negative for the analyte or protein in question.
- Warning: a strong positive result may safely be interpreted in as little as 2 minutes after sample addition. It is not safe, however, to interpret negative results prior to 5 minutes.
- Protect all components from hot or cold extremes of temperature when not in use.
- CAUTION: Tightly closed containers of soy extract, if left sitting for several hours, may ferment and cause the lid or container to burst. Dispose of extract when testing is complete.



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