ENVIROLOGIX™

QuickTox™ Kit for QuickScan Fumonisin Flex

Catalog AQ 311 BG *Part # 12210, 12239*

Matrices and Detection Ranges:

Matrix Group ID	Protocol	*Results reported in the range of:	Limit of Detection (LOD)*	Highest Approved Level*
FM MG1 - Corn	High Sensitivity 0.2 - 1.5 ppm	0 - 1.5 ppm	0.2 ppm	1.5 ppm
	Base Range 1.5 - 7 ppm	0 - 9 ppm	1.5 ppm	7.0 ppm
	High Positive 7 - 30 ppm	0 - 41 ppm	7.0 ppm	30 ppm

Do not assume accuracy for results reported below the protocol's LOD or above the protocol's highest approved level

Important Notes:

- Before testing, the enclosed Multi-Matrix Barcode Card (MMBC) must be scanned just once for each kit lot to upload information to the QuickScan
- Fold MMBC and scan only the MG1 barcode if you want QuickScan to skip the matrix selection and default to only MG1 matrices
- QuickScan Software Version 4.9.4 Update 1 or later is required
- DB6 Buffer is matched with specific Fumonisin Flex kit lot numbers. Be sure to use DB6 with the kit it is provided with. There is a "use with" label on the DB6 that will indicate the matching Fumonisin Flex Lot Number.

Table A on page 9 is provided as a Summary Guide for testing. More details for each step in the process are described below, and are important for achieving optimal, accurate results.

Contents of Kit:

- 50 QuickTox Strips packed in a moisture-resistant canister
- 50 clear Reaction tubes
- 50 blue Dilution tubes
- 100 pipette tips (1-200 µL)
- 50 pipette tips (100-1000 µL)
- DB6 Buffer, kit lot specific
- Multi-Matrix Barcode Card, kit lot specific

Items Not Provided:

- QuickScan System*
- Incubator base*
- Incubator block*
- Bunn grinder or equivalent
- 20-mesh screen
- Digital scale for weighing samples
- Extraction cups with lids* or other suitable vessels for sample extraction
- Graduated cylinder*
- Orbital/rotary shaker
- Pipette to deliver 200 µL*
- Pipette to deliver 50 µL*
- Pipette to deliver larger volumes (>200µL to 1 mL) for dilutions*
- Timer
- Scissors
- Distilled, deionized or bottled water

***Available as Accessories**

Available Accessories:

Item	*Catalog No.*	*Part #*
QuickScan™ System	ACC 131	10050 + 10198
50 Sample cups/lids *For extracting samples up to 30g; extracting larger samples requires different vessels.*	ACC 012-50	11224
Graduated cylinder (100mL)	ACC 068	11207
MiniPet pipette 200 µL (one/location free)	ACC 067	11206
MiniPet pipette 50 µL (one/location free)	ACC 051	11203
1 mL adjustable pipette	ACC 1303-PRO-1000	11964
Pipette tips for 1 mL pipette (50)	20-0127	12243
Incubator	ACC BSH300	12458

Intended Use

The QuickTox Kit for QuickScan Fumonisin Flex is designed to quickly provide quantitative results for the presence of total fumonisins.

- Limit of detection (LOD) = **0.20 ppm (high sensitivity protocol)**
- Assay range = 0.2 - 30 ppm, following three different protocols for the sub-ranges defined below.
  - 0.2 - 1.5 ppm ("High Sensitivity") - 1.5 - 7.0 ppm - 7.0 - 30 ppm ("High Positive")

How the Test Works

A composite sample is first collected, then extracted to solubilize any fumonisin present. Each sample should be ground to a fineness of 20 mesh and extracted using the specified extractant. This extract is further diluted for testing with the QuickTox Kit.

Each QuickTox Strip has an absorbent pad at each end. The protective tape with the arrow indicates which end of the strip to insert into the reaction vial. The sample extract travels up the membrane strip and is absorbed into the larger pad at the top of the strip. At the end of the test time, the strip is cut off at the top of the arrow tape, the bottom pads are discarded, and the strip is inserted into the QuickScan reader to obtain quantitative results.

Assay Preparation

Table A on page 9 is provided as a Summary Guide for testing. More details for each step in the process are described below, and are important for achieving optimal, accurate results.

Preparation of the Sample

Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. Make sure all reagents including samples, strips, buffer, and sample extractant are at room temperature and ready for use before starting the assay. The sample extract should be tested shortly after dilution with buffer.

Determine number and size of sub-samples and weigh out

1. Collect a composite sample according to your own sampling plan or USDA/GIPSA guidelines. Consult USDA/GIPSA reference documents to help design a plan that fits your needs.
2. Grind samples using a Bunn grinder or mill which provides a sample such that ≥95% passes through a 20-mesh sieve. Mix ground material thoroughly before sub-sampling.
3. Weigh samples into containers that will allow enough head room for the liquid to move forcefully when shaken vigorously.

Extract samples

1. Consult the Summary Guide Table A to determine the volume and type of Extractant that has been validated for the matrix. To calculate the volume of liquid to add, multiply the sample weight (in grams) x ratio (in milliliters, mLs)

   For example, 20 grams x 5 = 100 mL (water) to add to corn
2. Make sure the grain is completely wet, and then mix thoroughly as stated in the table. Liquid should be moving forcefully through the matrix to extract the fumonisin.
3. The order of addition has been optimized. Please follow this order.
4. Samples that are not thoroughly mixed and fully wetted may adversely affect test results due to inconsistent extraction.

Clarify extracts (again, adhere to the Summary Guide table for optimal performance)

1. Settling: Allow the sample to sit undisturbed until a top layer forms that can easily be pipetted. This top layer is the extract that will be used in the testing.

Protocol Selection Relative To Your Level(s) of Interest:

If your Level of Interest falls within the range of a single protocol, run only that protocol. If your level of interest spans the full quantitation range (0.2-30 ppm); it is recommended that you start with the Base Range followed by either the High Sensitivity or High Positive protocol depending on the results—this run order will minimize the time and number of strips required to get to the final result.

Protocol	*Results reported in the range of:	Limit of Detection (LOD)*	Highest Approved Level*	Sample Dilution 1	Sample Dilution 2	Transfer run volume to a clear Reaction tube and add to Incubator
High Sensitivity 0.2 - 1.5 ppm	0 - 1.5 ppm	0.2 ppm	1.5 ppm	350 µL DB6 buffer + 50 µL extract in blue Dilution tube	NA	Transfer 200 µL into clear Reaction tube
Base Range 1.5 - 7 ppm	0 - 9 ppm	1.5 ppm	7.0 ppm	2.5 mL DB6 buffer + 50 µL extract in blue Dilution tube	NA	Transfer 200 µL into clear Reaction tube
High Positive 7 - 30 ppm	0 - 41 ppm	7.0 ppm	30 ppm	2.5 mL DB6 buffer + 50 µL extract in blue Dilution tube	150 µL DB6 buffer + 50 µL Sample Dilution 1 in clear Reaction tube	

Do not assume accuracy for results reported below the protocol's LOD or above the protocol's highest approved level

Refer to Table A on p. 9 for the complete extraction and run instructions.

Add reagents to the blue Dilution Tube, followed by transfer to the clear Reaction Tube.

Reference Table A for protocol-specific dilutions based on the quantitation level desired.

1. **Take care not to contaminate the DB6 Buffer**. Keep Buffer covered when not in use, and use a new pipette tip for each test. **Please note**: DB6 Buffer is matched with specific Fumonisin Flex kit lot numbers; be sure to use the DB6 that is provided with the kit (do not mix and match buffers with different kit lots). There is a "use with" label on the DB6 that will indicate the matching Fumonisin Flex lot number.
2. Follow Table A instructions for Buffer and extract order of addition.
3. Use three pipette tips (large tip for Buffer, small tip for extract and another small tip to transfer the mixture to the Reaction tube) for each sample. *Retain the large pipette tip after buffer addition to be used for mixing purposes.
4. While adding the extract to the buffer in the Dilution Tube make sure to rinse the small tip by drawing it up and down a few times.
5. Mix Buffer and sample extract thoroughly by drawing the liquids up and down in the pipette tip (always use the larger volume pipette for this purpose). Samples that are not thoroughly mixed and/or accurately pipetted will adversely affect test results.
6. Transfer 200 µL of the diluted sample to the Reaction Tube.
7. Use a new Dilution Tube and Reaction Tube for each sample.
8. Follow the instructions under How to Run.

How to Run the QuickTox Strip Test

A minimum of 10 minutes before testing is to start, turn on the incubator and set to 22°C (follow manufacturer's instructions for setting temperature); ensure that the temperature display has stabilized and indicates "OK" before starting the assay. If testing is planned throughout the day it recommended to turn the incubator on in the morning and leave it on throughout the day.

1. Allow refrigerated canisters to come to room temperature before opening.
2. Add the reaction tube containing the diluted sample to the incubator (be sure it has reached 22°C). If the temperature of the testing environment is unknown or outside of the range of 20-24°C (68-75°F), **allow the sample to acclimate in the incubator for 2 minutes before proceeding.**
3. Remove the QuickTox Strips to be used. Avoid bending the strips. Reseal the canister immediately.
4. Place the strip into the reaction tube containing the Buffer and sample extract. The arrow tape on the end of the strip should point into the reaction tube.
5. Allow the strip to develop for the time noted in Table A (e.g., 5 minutes for corn).
6. Immediately cut off and discard the bottom section of the strip covered by the arrow tape. Insert strip into the QuickScan reader for quantitation.

Use of the QuickScan System

Detailed instructions for use of the QuickScan System are supplied with each unit, and can also be found at http://www.envirologix.com/support/quickscan. QuickScan Software Version 4.9.0 Update 1 or later is required and the lot-specific Multi-Matrix Barcode Card must be scanned into the system prior to testing.

In summary, a strip is inserted face down in the carrier with the barcoded end closest to the handle. The carrier is inserted into the reader and the strips are read by touching or clicking on the "Read Test" area of the screen. If the "Select Matrix Group" screen appears, select the group that displays the matrix run for each device. Refer to the Summary Guide Table A to determine if an alternative selection under the Dilution Tab is required based on the protocol that was run. Results are then recorded in an electronic worksheet, allowing each user to report and track data easily.

Based on the protocol run, ensure the appropriate selection is made under the Dilution tab on the results screen.

	Protocol Run		
	High Sensitivity 0.2 - 1.5 ppm	 1.5 - 7 ppm	High Positive 7 - 30 ppm
Dilution tab drop down menu selection	1:1 (this is the software default)	1:A (this must be selected)	1:B (this must be selected)

Kit Storage

This QuickTox Kit should be stored refrigerated. Note the shelf life on the kit box. Prolonged exposure to high temperatures may adversely affect the test results. Do not open the desiccated canister until ready to use the strips.

Cross-reactivity

The following mycotoxins have been tested with this kit and no false positive results occurred at the 200 ppm level: Aflatoxin B1, DON (deoxynivalenol), Ochratoxin A, Zearalenone.

Precautions and Notes

- Strips must be read wet promptly at the specified time for the matrix run to ensure accurate results.
- This product is currently not applicable for use in testing any other crops beyond those specified in this Product Insert.
- The corn assay is calibrated against samples with Fumonisin levels determined by a 3rd party using UHPLC/MS/MS with 13C isotopic internal Fumonisin standards (Biopure ILM003, ILM004 and ILM005, Romer Labs). Performance in other sample matrices has been validated using fortified samples.
- As with all screening tests, it is recommended that results be confirmed by an alternate method when necessary.
- The assay has been optimized for use with the protocols provided in the kit. Deviation from these protocols may invalidate the results of the test. Room-temperature components, proper and thorough mixing, accurate pipetting, and using the correct corresponding DB6 Buffer provided in the kit are essential to accurate results.
- The results generated through the proper use of this diagnostic tool reflect the condition of the working sample directly tested. Extrapolation as to the condition of the originating lot, from which the working sample was derived, should be based on sound sampling procedures and statistical calculations which address random sampling effects, non-random sampling effects and assay system uncertainty. A negative result obtained when properly testing the working sample does not necessarily mean the originating lot is entirely negative for the analyte in question.
- Protect all components from hot or cold extremes of temperature when not in use. Do not leave in direct sunlight or in vehicle.
- Observe any applicable regulations when disposing of samples and extracts.

For Technical Support Contact Us At:

EnviroLogix
500 Riverside Industrial Parkway
Portland, ME 04103-1486 USA
Tel: (207) 797-0300
Toll Free: 866-408-4597
Fax: (207) 797-7533

e-mail: ***info@envirologix.com***

website:
www.envirologix.com

LIMITED WARRANTY

EnviroLogix Inc. ("EnviroLogix") warrants the products sold hereunder ("the Products") against defects in materials and workmanship when used in accordance with the applicable instructions for a period not to extend beyond a product's printed expiration date. If the Products do not conform to this Limited Warranty and the customer notifies EnviroLogix in writing of such defects during the warranty period, including an offer by the customer to return the Products to EnviroLogix for evaluation, EnviroLogix will repair or replace, at its option, any product or part thereof that proves defective in materials or workmanship within the warranty period.

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This Limited Warranty states the entire obligation of EnviroLogix with respect to the Products. If any part of this Limited Warranty is determined to be void or illegal, the remainder shall remain in full force and effect.

License

EnviroLogix has developed this kit using proprietary reagents.

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Material Safety Data Sheet
According to OSHA 29CFR 1910.1200

SECTION 1. Identification of the substance/mixture and of the company/undertaking

1.1 Product identifier
Trade name: **DB 6 Dilution Buffer**
Part number 11151 (KR-268)

1.2 Relevant identified uses of the substance or mixture and uses advised against application of the substance / the preparation : Laboratory chemicals; kit component. Not to be used for purposes other than those specified in product literature.

1.3 Details of the supplier of the safety data sheet
Manufacturer/Supplier: EnviroLogix Inc., 500 Riverside Industrial Pkwy, Portland ME 04103, USA Phone: (207) 797-0300

1.4 Emergency telephone number: (207) 797-0300 Technical Service

SECTION 2. Hazards identification

2.1 Classification of the substance or mixture
Classification according to 29CFR 1910.1200: Not Classified

2.2 Label elements
Labeling according to 29CFR 1910.1200
Pictogram: None
Signal word: None
Hazard Statements: None

2.3 Other Statements: None

SECTION 3. Composition/information on ingredients

3.2 Mixture

Chemical name	CAS No	EC No	Classification According to 29CFR 1910.1200	Amount (%)
Sodium Tetraborate Decahydrate	1303-96-4	215-540-4	H360 Rep 1B	1 – 3 %

SECTION 4. First aid measures

4.1 Description of first aid measures
After inhalation : In case of inhalation. Remove to fresh air. If not breathing give artificial respiration. Get medical attention immediately.
After skin contact : In case of skin contact. Remove contaminated clothing and shoes immediately. Wash affected area with mild soap or detergent for at least 10 minutes or until no evidence of chemical remains.
After eye contact : In case of eye contact, immediately flush eyes with plenty of water for at least 15 minutes. Lifting eyelids occasionally, until no evidence of chemical remains. Get medical attention immediately
After swallowing : In case of ingestion. DO NOT Induce vomiting unless directed to do so by medical personnel. Never give anything by mouth to an unconscious person. Call a physician immediately.

4.2 Most important symptoms and effects, both acute And delayed: None

4.3 Indication of any immediate medical attention and special treatment needed: None

SECTION 5. Firefighting measures

5.1 Extinguishing media: CO_2, extinguishing powder or water spray. Fight larger fires with water spray or alcohol resistant foam.

5.2 Special hazards arising from the substance or mixture: None

5.3 Advice for firefighters: Wear protective gear appropriate for fire conditions including respiratory protective gear.

SECTION 6. Accidental release measures

6.1 Personal precautions, protective equipment and emergency procedures: In the case of spilled mixture wear gloves to prevent skin contact. In the case of a large spill, additional protection is recommended.

6.2 Environmental precautions: Do not discharge mixture to sewer system or waterways.

6.3 Methods and material for containment and cleanup: Absorb in paper towel or suitable absorbent for larger spills and discard in appropriate waste. Clean with water afterwards.

6.4 References to other sections: For safe handling refer to Section 7. For information on PPE refer to Section 8. For disposal refer to Section 13

SECTION 7. Handling and storage

7.1 Precautions for safe handling: Practice good chemical hygiene when handling. Avoid contact with eyes, skin, and clothing.

7.2 Conditions for safe storage, including any Incompatibilities: Store in tightly closed, non-metal container, in a corrosive compatible area. Prevent direct sunlight and heat. Store in well aired storage rooms.

7.3 Specific end use(s): Apart from the uses mentioned in section 1.2, no other specific uses are stipulated

SECTION 8. Exposure controls/personal protection

8.1 Exposure limits:
Components with limit values that require monitoring at the workplace:

	EH40/2005	OSHA
Sodium Tetraborate Decahydrate	8 Hr TWA = 5mg/m³	8 Hr TWA = 10 mg/m³

8.2 Exposure Controls:

8.2.1 Engineering controls: Facilities using this mixture should be equipped with an eyewash and safety shower. Use general or local exhaust ventilation to keep airborne concentrations below permissible exposure limits.

8.2.2 General protective and hygienic measures: The usual precautionary measures should be adhered to when handling chemicals.

Eye Protection: Safety glasses with side shields, goggles. Use equipment for eye protection tested and approved under appropriate government standards such as NIOSH (US) or EN 166 (EU). Eye and face protection regulations are described by OSHA (US) in 29CFR1910.133. Do not wear contact lenses when working with chemicals

Hand Protection: Handle with gloves. Gloves must be inspected prior to use. Use proper glove removal technique (without touching glove's outer surface) to avoid skin contact with this product. Dispose of contaminated gloves after use in accordance with applicable laws and good laboratory practices. Wash and dry hands. The selected protective gloves have to satisfy the specifications of EU Directive 89/686/EEC and the standard EN 374 derived from it.

Breathing Equipment: Appropriate respiratory protection should be determined according to local conditions using risk analysis protocols. An approved disposable air purifying particulate respirator may be used as a backup to engineering controls. Always use respirators and components tested and approved under appropriate government standards such as NIOSH (US) or CEN (EU).

8.2.3 Environmental exposure controls: Contain spills, do not allow into environment

SECTION 9. Physical and chemical properties

9.1 Information on basic physical and chemical properties:

a) Appearance:	Clear liquid, colorless to slight yellow.
b) Odor:	None
c) Odor Threshold:	No Data Available
d) pH:	8.6
e) Melting point/freezing point:	No Data Available
f) Boiling point/Boiling range:	No Data Available.
g) Flash point:	Not applicable.
h) Evaporation rate:	No Data Available
i) Flammability (solid, gaseous):	No Data Available
j) Upper/lower flammability or explosive limits:	No Data Available
k) Vapor pressure:	No Data Available
l) Vapor density	No Data Available
m) Relative density:	No Data Available
n) Solubility(ies):	Fully miscible, water.
o) Partition Coefficient: n-Octanol/water:	No Data Available
p) Auto-ignition temperature:	No Data Available
q) Decomposition temperature:	No Data Available
r) Viscosity:	No Data Available
s) Explosive properties:	No Data Available.
t) Oxidizing properties:	No Data Available

9.2 Other information No further relevant information available.

SECTION 10. Stability and reactivity

10.1 Reactivity: No data available

10.2 Chemical Stability: Stable under normal temperatures and pressures.

10.3 Possibility of hazardous reactions: Under normal conditions of storage and use, hazardous reactions will not occur.

10 4 Conditions to avoid: No specific data

10.5 Incompatible materials: No Data Available.

10.6 Hazardous decomposition products: Under normal conditions of storage and use, hazardous decompositions products should not be produced.

SECTION 11. Toxicological information

Information on Toxicological Effects
Acute effects (toxicity tests): No Data Available

Sensitization: No sensitizing effects known

CMR (carcinogenicity, mutagenicity and toxicity for reproduction) effects: No CMR effects

Additional toxicological information: No Additional Information

SECTION 12. Ecological information

12.1 Toxicity: No Data Available

12.2 Persistence and degradability : No Data Available

12.3 Bio accumulative potential: No Data Available

12.4 Mobility in soil : No Data Available

12.5 Results of PBT and vPvB assessment: Not available as a chemical safety assessment, not required/not conducted.

12.6 Other adverse effects: No Data Available

SECTION 13. Disposal considerations

Waste treatment methods: Contact a licensed professional waste disposal service to dispose of this material. Disposal of surplus or waste solutions must be in accordance with applicable local, state, and national laws and regulations.

SECTION 14. Transport information

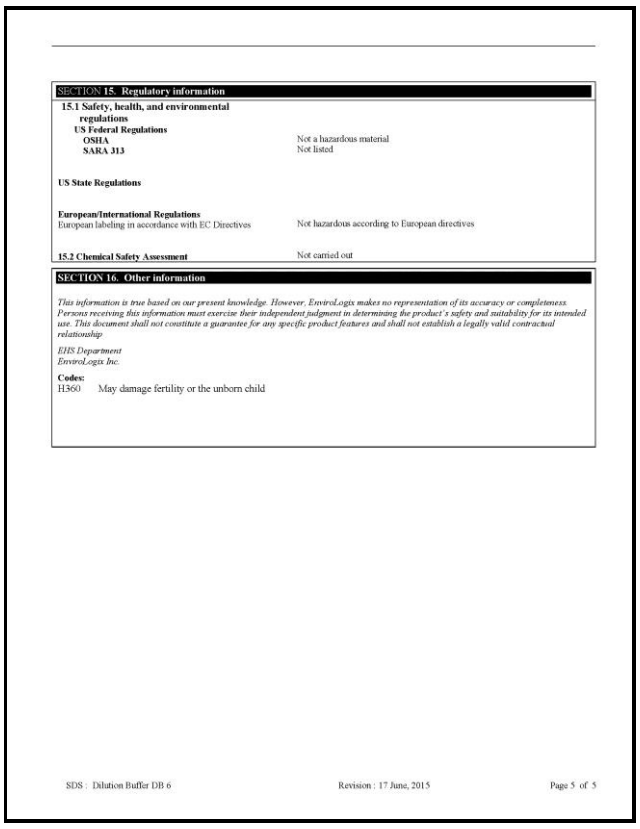
14.1 UN-Number DOT, ADR, ADN, IMDG, IATA : Not Hazardous for Transport
14.2 UN proper shipping name DOT, ADR, ADN, IMDG, IATA : Not Hazardous for Transport

14.3 Transport hazard class(es) DOT, ADR, ADN, IMDG, IATA): Not Hazardous for Transport
14.4 Packing group (DOT, ADR, IMDG, IATA): Not Hazardous for Transport

14.5 Environmental hazards No environmental hazard.

14.6 Special precautions for user : None

14.7 Transport in bulk according to Annex II of MARPOL73/78 and the IBC code: No information available.



SECTION 15. Regulatory information

15.1 Safety, health, and environmental regulations

US Federal Regulations

OSHA	Not a hazardous material
SARA 313	Not listed

US State Regulations

European/International Regulations

European labeling in accordance with EC Directives	Not hazardous according to European directives
15.2 Chemical Safety Assessment	Not carried out

SECTION 16. Other information

This information is true based on our present knowledge. However, EnviroLogix makes no representation of its accuracy or completeness. Persons receiving this information must exercise their independent judgment in determining the product's safety and suitability for its intended use. This document shall not constitute a guarantee for any specific product features and shall not establish a legally valid contractual relationship.

EHS Department
EnviroLogix Inc.

Codes:
H360 May damage fertility or the unborn child

SDS : Dilution Buffer DB 6 Revision : 17 June, 2015 Page 5 of 5

Table A: Summary Guide for Approved Matrices

Approved Matrices (associated assay range)	Matrix Group	Add Grain to Vessel First	Add Extractant Second	Fully wet sample, then mix	Clarify	Run the Base Range protocol first followed by either the High Positive or High Sensitivity protocols if necessary[#]	Pre-Mix sample in blue Dilution Tube followed by transfer to clear Reaction Tube	Add Reaction Tube to Incubator set at 22°C	Add strip for	Read in QuickScan: Dilution tab on the result page should display
Corn	FM MG1	20g to 50g	5x vol water* 5 mL per gram of sample, e.g. 250 mL to a 50g sample	1 minute highest speed on shaker table, *or* 2 minutes vigorously by hand	Settle	1.5 to 7.0 ppm (Base Range)	**Pre-Mix** 2.5 mL buffer + 50 µL extract† **Transfer** 200 µL	Acclimate tube for 2 min^	5 min.	1:A (this must be selected)
						7.0 to 30 ppm (High Positive)	**Transfer** 150 µL buffer + 50 µL of the Pre-Mix extract from the 1.5 - 7 ppm protocol, Mix	Acclimate tube for 2 min^	5 min.	1:B (this must be selected)
						0.2 to 1.5 ppm (High Sensitivity)	**Pre-Mix** 375 µL buffer + 50 µL extract **Transfer** 200 µL	Acclimate tube for 2 min^	5 min	1:1 (this is the software default)

Notes:

* Use distilled, deionized, or flat (non-carbonated) bottled water.

^ The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20-24°C (68-75°F)

† Retain this Pre-Mix extract in case High Positive testing is necessary

[#] If your Level of Interest falls within a single protocol range, run only that protocol (see Instructions and table on p. 3-4)